Supplementary Information for

Spinal Cord Injury Affects Gene Expression of Transmembrane Proteins in Tissue and Release of Extracellular Vesicle in Blood: In Silico and In Vivo Analysis

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Fig.S1: DNA microarray data selection. The microarray datasets related to spinal cord injury are selected according to the type of sample, stage of injury, and region of injury.

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Table S1: Sample selection in each dataset					
Dataset	GSE464	GSE2599	GSE45006	GSE46988	
Phase					
Acute	4 samples intact	-	4 samples sham injury	4 samples intact	
	4 samples 7d PI		4 samples 7d PI	4 samples 7d PI	
Subacute	4 samples intact	3 samples control	4 samples sham injury	-	
	4 samples 28d PI	3 samples 35d PI	4 samples 14d PI		

From each dataset in both injury phases, a control group and a post-injury group were selected, and the number of samples in each group is given in the table.

Table S2: Animal procedures groups					
Groups	Condition	State	Samples		
1	Control	Intact	3		
2	Sham	Laminectomy (7 day)	3		
3	Sham	Laminectomy (14 day)	3		
4	SCI	Acute (7 day)	3		
5	SCI	Subacute (14 day)	3		

BBB score	After injury	7 d PI	14 d PI
Rat number			
1	0	1.5	-
2	0.5	4	-
3	0.5	2	-
4	0.5	5	9.5
5	0.5	1.5	2
6	1	5	8.5

BBB; Basso, Beattie, and Bresnahan, d; Day, and PI; Post Injury.



Fig.S2: Timeline of the experiment. As the picture depicts, SCI surgery is considered as day 0. One week after that acute group and after 2 weeks subacute group have sacrificed. Caring and other assessments have performed in the following weeks. SCI; Spinal cord injury, BBB; Basso, beattie, and bresnahan, and RT-PCR; Real time polymerase chain reaction.



Fig.S3: SDS-PAGE was used to obtain high-resolution separation of complex mixtures of proteins.



Fig.54: Overlapped DEGs in the acute and subacute phases. In acute and subacute phases, differential expressed genes were shared across the three datasets, and genes that were common to at least two datasets were selected. DEGs; Differentially Expressed Genes.

Table 54: Primers sequences						
Gene name	Primer sequences (5 -3)	Accession number	Annealing temperature (°C)	Amplicon size (bp)	IM (°C)	
Enpp3	F: GTATCCAGAGTCGCACGGCA	NM_019370	60	197	62	
	R: AGGAGCCATTGACAGCCACA					
Cd63	F: GCATCAAGAAGCGTCGGGGA	XM_008765024	59	194	63	
	R: ACAGCGAGCCAGCAGTAGTC					
Cxcr4	F: GGCCGTCTATGTGGGTGTCT	NM_022205	60	191	62	
	R: AGGACAGGATGACGATGCCC					
Kcnal	F: CCCGAGAAGGAGTACCAGCG	XM_039107020	60	174	62	
	R: AATGGTGCCCGTGAAGTCCT					
Scn1a	F: TTCCCCGAGACGCAATAGCA	NM_030875	58	196	61	
	R: GGAGGATCTGCTGGTTTGGC					
Nrg1	F: CACCTTTCCCTCTTCGGGCT	NM_001271118	59	154	61	
	R: AGGAAGCAGCGTGGACTCG					
Grm1	F: ACGGCGAGAGTGGAATGGATGC	NM_001114330	60	169	65	
	R: TCGCAGAAGCAGACCACAACCC					

TM; Melting temperature.



Fig.S5: Melting curve for the quantitative real-time polymerase chain reaction (PCR) of RNA samples treated with DNase I and reverse transcribed to cDNA.