Nitric Oxide Levels and Total Antioxidant Capacity in The Seminal Plasma of Infertile Smoking Men

Yousefreza Yousefniapasha, M.D.¹, Gholamali Jorsaraei, Ph.D.¹, Maryam Gholinezhadchari, M.Sc.¹*, Soleiman Mahjoub, Ph.D.¹,², Mahmoud Hajiahmadi, Ph.D.³, Mehrdad Farsi, Ph.D.¹

Fatemeh-zahra Infertility and Reproductive Health Research Center, Babol University of Medical Sciences, Babol, Iran
Department of Biochemistry and Biophysics, Faculty of Medicine, Babol University of Medical Sciences, Babol, Iran
Infectious Diseases and Tropical Medicine Research Center, Babol University of Medical Sciences, Babol, Iran

*Corresponding Address: P.O.Box: 47135-547, Fatemeh-zahra Infertility and Reproductive Health Research Center, Babol University of Medical Sciences, Babol, Iran Email: Mgh.chari@gmail.com

Received: 18/Sep/2013, Accepted: 10/Jan/2014

Abstract

Objective: Cigarette is a rich source of oxidants and reactive nitrogen species. Nitric oxide (NO) in high concentration has deleterious effects on human sperm function. Antioxidant defense system in seminal plasma protects spermatozoa from the attack of reactive oxygen metabolites. The purpose of this study was to determine the correlation between cigarette smoking with the NO levels and the total antioxidant capacity (TAC) of the seminal plasma in infertile smoker men and to compare severity of oxidative stress (OS) in them with fertile and infertile non-smoking men.

Materials and Methods: In this cross sectional study, a total of 95 male participants attended the Infertility Clinic of the Fatehmeh-zahra Hospital in Babol, Mazandaran Province, Iran, between 2010 and 2011. They were divided into three groups: I. fertile non-smokers (F.ns; n=32), II. infertile non-smokers (IF.ns; n=30) and III. infertile smokers (IF.s; n=33) according to semen analysis World Health Organization guidelines (WHO, 2001) and smoking data. TAC concentration and NO levels of seminal plasma were measured using the ferric reducing ability of plasma (FRAP) method and the Griess reagent, respectively.

Results: Standard sperm parameters were significantly higher in the fertile group than those in the infertile groups, but these differences between the IF.ns and IF.s were not statistically significant. The mean TAC in the seminal plasma was higher in the F.ns>IF.ns>IF.s, respectively, but these differences were not statistically significant among three groups (p= 0.096). In contrast, the mean NO level in the seminal plasma was lower in the F.ns<IF.ns< IF.s, respectively. These differences were statistically significant among the three groups (p= 0.018).

Conclusion: It argued that the increased NO levels associated with smoking might exceed the capacity of antioxidant defense system, leading to increased oxidative damage of seminal plasma and decreased fertility in men.

Keywords: Smoking, Nitric Oxide, TAC, Male Infertility

Cell Journal (Yakhteh), Vol 17, No 1, Spring 2015, Pages: 129-136

Citation: Yousefniapasha YR, Jorsaraei GhA, Gholinezhadchari M, Mahjoub S, Hajiahmadi M, Farsi M. Nitric oxide levels and total antioxidant capacity in the seminal plasma of infertile smoking men. Cell J. 2015; 17(1): 129-136.

Introduction

Cigarettes contain a wide category of xenobiotics, including oxygen free radicals and reactive nitrogen species (RNS) (1). It has been shown that the enlarged production of reactive oxygen species (ROS) related to smoking could increase the capacity of oxidant defense system, resulting in redoubled aerobic injury of seminal plasma (2,

3). Smoking-tobacco contains nearly 4000 harmful compounds like alkaloids, nitrosamines, nicotine and hydroxycotinine that many of these substances generate ROS and RNS (4). Smoking has been shown to be associated with a 48 and 107% increase in seminal leukocyte and ROS concentrations, respectively (5). Among the body cells, spermatozoa due to small amounts of cytoplasm en-

zymes are very sensitive to oxidative damages and are unable to repair them (6). On the other hand, the plasma membrane of spermatozoa possesses high polyunsaturated fatty acids (PUFA), which make them susceptible to ROS by inducing lipid peroxidation (LPO) (6, 7). A physiological level of ROS during normal aerobic metabolism in spermatozoa is essential for maturation, capacitation, acrosomal reactions, and spermatozoan-oocyte fusion as well as for the general function or viability of spermatozoa (8, 9). However, the production of excessive levels of ROS can be potentially toxic for sperm function (6, 7). High levels of ROS may also affect the sperm axoneme, inhibit mitochondrial function and disturb the synthesis of nucleic acids (DNA and RNA) and proteins (10). ROS decreases motility of spermatozoa due to damages to the flagellum and axonemal structures of their tails (11).

Furthermore, it has been shown that ROS can damage DNA by causing base modification, DNA strand breaks, DNA cross-links, and chromosomal rearrangements (9, 12). Immature or abnormal spermatozoa and leukocytes are two major sources of production of free radicals in semen (2, 6). The most common ROS with significant effects in reproductive biology are as follows: superoxide anion (O, -), hydrogen peroxide (H,O,), peroxyl radicals (ROO), and also the highly reactive hydroxyl radicals (OH⁻) (7). The nitrogen species derived from nitric oxide (NO•) and peroxynitrite anion (ONOO-) additionally seem to play a significant role within the reproduction and fertilization (7). NO is an oxygen free radical generated from the oxidation of L-arginine to L-citrulline by three isoforms of nicotinamide adenine dinucleotide phosphate (NADPH)-dependent NO synthases (NOS) (13). However, it has been additionally reported that formation of NO and ONOO- are caused by use of D-arginine in class spermatozoa mistreatment, cannot be a substrate for NOS. Under these conditions, NO production may be induced nonenzymatically through a H₂O₂ mediated attack on arginine (14) and also shows cytotoxic effects that are mediated indirectly through its interaction with O, and formation of ONOO, which decomposes to form OH and NO₂, respectively (15). NO features a twin impact on sperm performs, so under physiological conditions, NO plays a vital role in normal sperm production and motility. Low NO levels have been shown to increase sperm motility (16), capacitation (17) and zonapellucida spermbinding protein (18), whereas a number of studies have shown a negative effect of high NO levels in seminal plasma on human sperm motility (13, 19). Overall, these studies have suggested a relevant role of NO within the pathophysiology of spermatozoa. The oxidative damage induced by free radicals is commonly decreased by a mix of biological inhibitor systems as total antioxidant capacity (TAC) that contains both enzymatic and non-enzymatic reactions. Human seminal plasma comprises the following three main enzymatic antioxidants: superoxide dismutase (SOD), catalase, and glutathione peroxidase/glutathione reductase (GPX/GRD). Furthermore, there is a wide range of non-enzymatic antioxidants as follows: ascorbate, urate, vitamin E, pyruvate, glutathione, albumin, vitamin A, ubiquitol, taurine, and hypotaurine (11). A ntioxidants are the major defense system against oxidative stress (OS) induced by free radicals. However, OS is the result of an imbalance between the generation of ROS and the antioxidant system that would have harmful effects on human spermatozoa (6, 7, 9). This study aimed to estimate the effects of cigarette smoking on the increased NO levels and decreased TAC in seminal plasma and to compare severity of OS in the infertile smoker men with fertile and infertile non-smoking men.

Materials and Methods

Sample collection and semen analysis

In this cross sectional study, a total of 95 semen samples were provided by donors with the age range 24-38 years attending the Infertility Clinic of the Fatemeh-zahra Hospital in Babol, Mazandaran Province, Iran, between 2010 and 2011. The individuals with a significant medical history, signs of defective and rogenisation, abnormal testicular examinations, chromosomal disorders related to a fertility disorder, cryptorchidism, vasectomy, endocrine disorders, leukocytospermia, alcohol consumption and treated with antioxidant supplements were excluded from this study. The semen samples were collected into sterile container after anabstinence period of 2-3 days at the in vitro fertilization (IVF) center. Specimens were allowed to liquefy at 37°C for 30 minutes. The routing analysis of sperm parameters including volume, count, total count and motility was performed within 1 hour according to World Health Organization guidelines (WHO, 2001) using microscopic examination (20). Sperm morphology <14% were considered abnormal according to Kruger's criteria (21). Then, based on demographic information and medical history of the participants such as age and other disease at the time of sampling/spermogeram analysis, the subjects were classified into three groups including: i. fertile non-smokers (F.ns; n=32) who had normal semen, ii. infertile non-smokers (IF.ns; n=30) and iii. infertile smokers (IF.s; n=33). Both IF.s and IF.ns groups had a history of primary infertility at least for 1 year and showed abnormal semen analysis, while smokers had consumed 10-20 cigarettes per a day for at least 2 years.

Measurement of TAC using the ferric reducing ability of plasma (FRAP) method

The semen samples were centrifuged at 12000 g for 7 minutes and stored at -20°C until analyt sis. Seminal plasma was thawed at room temperature and diluted 1:10 v/v in distilled water. TAC was then evaluated using the FRAP method according to the method described by Benzie and Strain (22) with slight modifications. This method measures the ability of antioxidants of a sample to reduce ferric-tripyridyltriazine (Fe³⁺-TPTZ) to a ferrous form (Fe²⁺). In brief, the working FRAP reagent was prepared by mixing 300 mmol/L acetate buffer pH=3.6 (3.1 g sodium acetate×3H₂O (Merck, Germany) and 16 ml acetic acid in 1000 ml buffer solution) with 10 mmol/L 2,4,6,-tripyridyls-triazine (TPTZ, Merck, Germany), 40 mmol/l Hydrochloric acid (HCl; Merck, Germany), and 20 mmol/l FeCl₂×6H₂O (Sigma, America) in a 10:1:1 ratio, just before use. Then, 1.5 mL of the working FRAP reagent was aliquoted into a glass tube and warmed to 37°C for 5 minutes. Subsequently, 50 µL of plasma, 50 µL of distilled water (reagent-free), as well as 50 µL of each of the standard solutions (FeSO₄.7H₂O; 1000, 500, 250, and 125 µM, Sigma, America) were added to 1.5 mL FRAP reagent and heated in 37°C for 10 ming utes. Absorbance was measured at 593 nm using a spectrophotometer (UV1600, Germany). The final results were expressed as µM/L.

Measurement of NO using the Griess reagent method

The total stable oxidation products of NO metabolism (NO₂-/NO₃-) of seminal plasma were as-

sessed using a Griess reagent. The Griess reagent consists of sulfanilamide (SULF) and N-(1-Naphthyl) ethylenediaminedihydrochloride (NEDD) (23, 24). The frozen semen was allowed to thaw and to reach a temperature of 25°C that was fola lowed by being deproteinized by zinc sulfate solution (Sigma, America). The liquefied semen was then centrifuged at 12000 g for 10 minutes. Aliquots (300 µL) of the clear supernatant was mixed with Griess reagents including 300 µL SULF (2% w/v, Sigma, America) in 5% HCl and 300 μL NEDD (0.1% w/v, Sigma, America) in H₂O in a test tube, while for the reduction of nitrate to nitrite, 300 µL saturated solutions of vanadium (III) chloride (VCl₂; Sigma, America) in 1 M HCl was added and incubated for 2 hours at 30°C in the dark. Then, the absorbance of samples was measured at 540 nm against a blank containing the same concentrations of ingredients but no biological sample. Linear regression was used to determine NO concentration from standard curve of NaNO₂. The final results were expressed as µmol/L.

Statistical analysis

One-way analysis of variance (ANOVA) was used to compare NO and TAC levels as well as other parameters including age, volume, morphology and total sperm count among the groups. The data were presented as mean ± standard deviation (SD). Percentage motility and sperm count were not normally distributed; therefore, the differences among the groups were done using the Median test (25th and 75th percentiles). The odds ratios (OR) were presented with their 95% confidence intervals (CI) using the statistical package for the social sciences (SPSS; SPSS Inc., Chicago, IL, USA) version 18. In all cases, a p value of <0.05 was considered statistically significant.

Ethical considerations

This study was conducted with the approval of the Medical Research Ethics Committee of the Faculty of Babol University of Medical Sciences, Babol, Iran. An informed written consent was obtained from all subjects participating in the study.

Results

After studying thedemographic information and medical history, a total of 95 semen samples

were divided into three groups: F.ns (n=32), IF.ns (n=30), including 1 asthenoterato, 1 oligo, 2 astheno, 4 oligoterato, 15 terato and 7 oligoasthenoterato; and IF.s (n=33) including 4 asthenoterato, 4 oligoterato, 18 terato and 7 oligoasthenoterato.

Semen analysis

According to the present results, all sperm parameters, including count, motility and morphology, among the three groups of the F.ns, IF.ns and IF.s were strongly significant (p<0.0001, Table 1). But these differences in standard sperm variables between the IF.s and IF.ns were not statistically significant. Semen volume and the mean of age showed no significant difference among the three

groups (p=0.41 and p=0.56, respectively).

TAC and NO assays

Comparison of the TAC and NO levels is shown in table 2. The mean NO levels in the seminal plasma was lower in the F.ns<IF.ns< IF.s, respectively. Also, NO levels showed a significant difference among the three groups (p=0.018). In contrast, the mean TAC levels in the seminal plasma was higher in the F.ns>IF.ns> IF.s, respectively, but these differences were not statistically significant among three groups (p=0.096, Table 2). Moreover, TAC and NO levels showed significant differences between F.ns and IF.s groups (p=0.05 and p=0.01, respectively).

Table 1: Comparison of sperm parameters in semen samples among the three groups of fertile non-smoking, infertile smoking and non-smoking men

Variables	F.ns (n=32)	IF.ns (n=30)	IF.s (n=33)	P value			
	Mean ± SD	$Mean \pm SD$	$Mean \pm SD$				
Volume (ml)	3.79 ± 1.39	3.95 ± 1.92	3.43 ± 1.29	0.407			
Sperm morphology (%)	31.22 ± 14.21	7.40 ± 5.17	6.67 ± 4.30	< 0.0001			
Total spermcount (×106)	337.50 ± 140.90	164.83 ± 147.26	155.41 ± 148.72	< 0.0001			
	Median (25 th , 75 th)	Median (25 th , 75 th)	Median (25 th , 75 th)				
Sperm count (×10 ⁶ /ml)	100.0 (82.0, 100.0)	30.0 (10.0, 72.5)	40.0 (11.0, 70.0)	< 0.0001			
Sperm motility (%)	60.0 (60.0, 70.0)	50.0 (37.5, 50.0)	50.0 (40.0, 60.0)	< 0.0001			

F.ns; Fertile non-smokers, IF.ns; Infertile non-smokers, IF.s; Infertile smokers, SD; Standard deviation and n; Number. P< 0.05 was considered statistically significant. One-way analysis of variance (ANOVA) was used for comparisons of volume, morphology and total sperm count among groups. Median test was used for comparisons of sperm count and motility among groups.

Table 2: Comparison of TAC and NO concentration of the seminal plasma among the three groups of fertile non-smoking, infertile smoking and non-smoking men

Variables	F.ns (n=33)	IF.ns (n=30)	IF.s (n=32)	P value
TAC (µM/L)	2136.25 ± 722.24	1799.17 ± 729	1785.45 ± 718.180	0.096
NO (μmol/L)	5.26 ± 3.32	7.26 ± 5.53	8.61 ± 4.96	0.018

Values are expressed as mean ± SD. TAC; Total antioxidant capacity, NO; Nitric oxide, F.ns; Fertile non-smokers, IF.ns; Infertile non-smokers, IF.s; Infertile smokers, SD; Standard deviation and n; Number. P< 0.05 was considered statistically significantusing one-way ANOVA.

Discussion

The etiology of defective sperm function is closely associated with the over production of ROS. It is important to concentrate on these signaling molecules involved in essential sperm pathways such as capacitation and oxidative damage in spermatozoa (25). In this study, we investigated the relationship between cigarette smoking with NO and TAC levels of seminal plasma to show an increase in oxidative damage level and infertility rate in men. Fraga et al. (26) found the level of 8-hydroxy-2'-deoxyguanosine (8-OHdG), a marker of DNA oxidation, is 50% higher in smokers compared with non-smokers (p=0.005). Smoking may also result in chromatin abnormalities and/or endogenous DNA strand breaks in human spermatozoa (27). Several studies have showed that cigarette smoking has an effect on reduced sperm quality (count, motility and morphology), especially for those who are heavy smokers or have smoked for many years (5, 28-32). In a study from Denmark, of >2500 healthy male participating, a statistically significant inverse dose-response relationship was observed between current cigarette smoking and semen analysis (semen volume, total sperm count and percentage of sperm motility). Heavy smokers (>20 cigarettes per day) had ~19% lower mean sperm concentration and a 29% lower total sperm count than non-smokers (32). These studies have shown that the decreased sperm quality in smokers can be due to OS. Pasqualotto et al. (31) evaluated 522 F.ns and 367 fertile smokers in three groups based on the mode of smoking doses. They found a decreasing semen volume with an increasing number of cigarettes smoke, but no statistically significant differences were observed among the groups in terms of sperm concentration and motility. Studies have also shown that smokers are more susceptible to OS, leading to adecline in seminal plasma antioxidants such as vitamin E (26) and vitamin C (33). According to a study of 40 smoking and non-smoking male university students (22-25 years) in West Bengal, India, malondialdehyde (MDA) level and the lactate dehydrogenase (LDH) activity in serum and neutrophil were higher in smokers (n=20) than the non-smokers (n=20). The activities of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), and glutathione

reductase (GR), as well as glutathione (GSH) levels were significantly lower in smokers than the non-smokers in both serum and neutrophil (3). They suggested that the oxidative damage found in smokers could be the result of direct effects of oxidants in cigarette smoke on the decreased antioxidant status (3). High levels of LPO with concomitant depletion of antioxidants in seminal plasma were observed in an Indian population consisting of 100 smokers as compared to 100 non-smokers subjects (34). Moreover, enzymatic and non-enzymatic antioxidants activity was significantly higherin smokers than non-smokers. They have suggested that probably some of the harmful effects of smoking are mediated by free radicals, leading to presence of LPO and breakdown of antioxidant status in cigarette smoking. It suggests that NO may be involved in the etiology of defective sperm function, as well as has a bilateral role, being both a cytotoxic and a necessary molecule for normal sperm production and motility that depends on the alternative redox state and relative NO levels (16, 19). A significant linear negative correlation has been reported between NO levels in seminal plasma and sperm motility (13, 19, 35), morphology (19, 35, 36) sperm count (19) and viability (35) in infertile men. It has been reported that O₂, H₂O₂ and NO can activate target membranes in order to trigger the intracellular mechanisms involved in sperm capacitation. Catalase blocks NO-induced capacitation, but the mechanisms of action involving H₂O₂ are still unknown. NO may react with O₂ to form ONOO, leading to reaction with glutathione, cysteine, deoxyribose, and other thiols/thioethers (37). This can result in formation of reactive nitrating species in the presence of metal ions or complexes. The dual effects of NO depend on its concentration and reactions with ROS such as H₂O₂ (37, 38). The first mechanism, NO inducing sperm damage, is likely to be involved in inhibition of mitochondrial respiration and DNA synthesis using nitrosylation of heme in mitochondrial enzymes, aconitase, and glyceraldehyde phosphate dehydrogenase that results in depletion of adenosine triphosphateand the reduction of sperm motility (38). A study in Italy has provided evidence that NO concentration in the seminal plasma of the idiopathic as the nozoospermic infertile patients

is significantly higher than those in normozoospermic fertile subjects. Moreover, they showed a significant negative linear correlation between NO concentration and sperm motility, as well as other kinetic parameters of spermatozoa such as VCL and VSL (13). Therefore, they have suggested that the high NO levels and the consequent excessive exposure to OS have a potential pathogenetic role in human sperm motility (13). Sheikh et al. (39) showed higher levels of DNA damage and NO in infertile men compared with fertile men. On the other hand, TAC levels in infertile men were significantly lower than fertile men. Also, a negative correlation was observed between NO concentration and TAC levels in infertile cases (p<0.001; r=-0.293). Amiri et al. (35) demonstrated that mean NO level of seminal plasma in infertile men was significantly higher than fertile donors. Badade et al. (19) investigated seminal levels of NO and TAC in fertile and infertile men. In their study, seminal levels of NO were significantly higher (p<0.001), while TAC was significantly lower (p<0.001) in infertile patients than fertile men. In our previous study, a significant increase in MDA levels and decrease in the mean TAC in the seminal plasma were found in asthenoteratospermic smokers and non-smokers as compared to normozoospermic men, but no significant differences were observed between the smoking and non-smoking patients (30). In the present study, TAC concentration of seminal plasma was higher in F.ns than IF.ns and IF.s (F.ns>IF.ns>IF.s, respectively). But these differences were not statistically significant among the three groups (p=0.096). In contrast, seminal NO levels showed a significant difference among the groups (p=0.018), so it was higher in IF.s than IF.ns and also F.ns (IF. s>IF.ns>F.ns, respectively). However, the following limitations were applied in our study. Smoking data, as all the other variables in the present study, were collected through self-completed questionnaires, which may have introduced a risk of misclassification of the exposure variable. In addition, the participation rate was low in our study, which may be the reason that we were not able to detect the significant correlation between parameters. Although we included subjects who smoked a minimum of 10 cigarettes per day for at least 2 years, we

were unable to classify this group (IF.s) based on the number of cigarettes consumed per day and the duration of smoking in years and to examine the relationship between smoking and semen parameters in a dose-dependent fashion because of the subjective nature of the smoking history. The other limitation of our study may be the lack of information regarding the exact etiology of infertility or reproductive potential in these patients. Despite these limitations, our study shows that cigarette smoking and infertility in men are related with increased levels of seminal OS.

Conclusion

It is deduced from our results that infertile smoking men have higher levels of seminal OS than fertile and IF.ns. The correlation between cigarette smoking and high OS levels in the seminal plasma can be attributed in part to the associated increase in seminal NO levels and an impaired oxidant defense system. It suggests that smoking induces OS either with the increased levels of oxidants originating from the smoke such as NO or with the decreased levels of antioxidants in the seminal plasma, leading to a decrease in fertilizing potential of spermatozoa in infertile men.

Acknowledgments

This study, conducted in Department of Biochemistry and Biophysics, Fatemeh-zahra Infertility and Reproductive Health Research Center, Babol, Iran, was financially supported by Babol University of Medical Sciences. We express our appreciation to Dr. Evangeline Foronda for the English editing of the article. There is no conflict of interest in this study.

References

- Pryor WA. Biological effects of cigarette smoke, wood smoke, and the smoke from plastics: the use of electron spin resonance. Free Radic Biol Med. 1992; 13(6): 659-676.
- Lavranos G, Balla M, Tzortzopoulou A, Syriou V, Angelopoulou R. Investigating ROS sources in male infertility: a common end for numerous pathways. Reprod Toxicol. 2012; 34(3): 298-307.
- Mahapatra SK, Das S, Dey SK, Roy S. Smoking induced oxidative stress in serum and neutrophil of the university students. Al Ameen J Med Sci. 2008; 1(1): 20-31.

- Traber MG, van der Vliet A, Reznick AZ, Cross CE. Tobacco-related diseases. Is there a role for antioxidant micronutrient supplementation?. Clin Chest Med. 2000; 21(1): 173-187.
- Saleh RA, Agarwal A, Sharma RK, Nelson DR, Thomas AJ Jr. Effect of cigarette smoking on levels of seminal oxidative stress in infertile men: a prospective study. Fertil Steril. 2002; 78(3): 491-499.
- Sikka SC, Rajasekaran M, Hellstrom WJ. Role of oxidative stress and antioxidants in male infertility. J Androl. 1995; 16(6): 464-468.
- Maneesh M, Jayalekshmi H. Role of reactive oxygen species and antioxidants on pathophysiology of male reproduction. Indian J Clin Biochem. 2006; 21(2): 80-89.
- Baumber J, Ball BA, Gravance CG, Medina V, Davies-Morel MC. The effect of reactive oxygen species on equine sperm motility, viability, acrosomal integrity, mitochondrial membrane potential, and membrane lipid peroxidation. J Androl. 2000; 21(6): 895-902.
- Cocuzza M, Sikka SC, Athayde KS, Agarwal A. Clinical relevance of oxidative stress and sperm chromatin damage in male infertility: an evidence based analysis. Int Braz J Urol. 2007; 33(5): 603-621.
- de Lamirande E, Gagnon C. Impact of reactive oxygen species on spermatozoa: a balancing act between beneficial and detrimental effects. Hum Reprod. 1995; 10 Suppl 1: 15-21.
- Agarwal A, Makker K, Sharma R. Clinical relevance of oxidative stress in male factor infertility: an update. Am J Reprod Immunol. 2008; 59(1): 2-11.
- Kemal Duru N, Morshedi M, Oehninger S. Effects of hydrogen peroxide on DNA and plasma membrane integrity of human spermatozoa. Fertil Steril. 2000; 74(6): 1200-1207.
- Balercia G, Moretti S, Vignini A, Magagnini M, Mantero F, Boscaro M, et al. Role of nitric oxide concentrations on human sperm motility. J Androl. 2004; 25(2): 245-249.
- Aitken RJ, Ryan AL, Baker MA, McLaughlin EA. Redox activity associated with the maturation and capacitation of mammalian spermatozoa. Free Radic Biol Med. 2004; 36(8): 994-1010.
- Beckman JS, Beckman TW, Chen J, Marshall PA, Freeman BA. Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. Proc Natl Acad Sci USA. 1990; 87(4): 1620-1624.
- Herrero MB, de Lamirande E, Gagnon C. Tyrosine nitration in human spermatozoa: a physiological function of peroxynitrite, the reaction product of nitric oxide and superoxide. Mol Hum Reprod. 2001; 7(10): 913-921.
- Zini A, De Lamirande E, Gagnon C. Low levels of nitric oxide promote human sperm capacitation in vitro. J Androl. 1995; 16(5): 424-431.
- Sengoku K, Tamate K, Yoshida T, Takaoka Y, Miyamoto T, Ishikawa M. Effects of low concentrations of nitric oxide on the zona pellucida binding ability of human spermatozoa. Fertil Steril. 1998; 69(3): 522-527.
- Badade ZG, More KM, Narshetty JG, Badade VZ, Yadav BK. Human seminal oxidative stress: correlation with antioxidants and sperm quality parameters. Ann Biol Res. 2011; 2(5): 351-359.
- World Health Organization. Laboratory manual of the WHO for the examination of human semen and sperm-

- cervical mucus interaction. Ann 1st Super Sanita. 2001; 37(1): I-XII, 1-123.
- 21. Kruger TF, Menkveld R, Stander FS, Lombard CJ, Van der Merwe JP, van Zyl JA, et al. Sperm morphologic features as a prognostic factor in in vitro fertilization. Fertil Steril. 1986; 46(6): 1118-1123.
- Benzie IF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. Anal Biochem. 1996; 239(1): 70-76.
- 23. Moshage H, Kok B, Huizenga JR, Jansen PL. Nitrite and nitrate determinations in plasma: a critical evaluation. Clin Chem. 1995; 41(6 Pt 1): 892-896.
- Mehraban D, Ansari M, Keyhan H, Sedighi Gilani M, Naderi G, Esfehani F. Comparison of nitric oxide concentration in seminal fluid between infertile patients with and without varicocele and normal fertile men. Urol J. 2005; 2(2): 106-110.
- Baker MA, Krutskikh A, Aitken RJ. Biochemical entities involved in reactive oxygen species generation by human spermatozoa. Protoplasma. 2003; 221(1-2): 145-151.
- Fraga CG, Motchnik PA, Wyrobek AJ, Rempel DM, Ames BN. Smoking and low antioxidant levels increase oxidative damage to sperm DNA. Mutat Res. 1996; 351(2): 199-203.
- Potts RJ, Newbury CJ, Smith G, Notarianni LJ, Jefferies TM. Sperm chromatin damage associated with male smoking. Mutat Res. 1999; 423(1-2): 103-111.
- 28. Liu RZ, Gao JC, Zhang HG, Wang RX, Zhang ZH, Liu XY. Seminal plasma zinc level may be associated with the effect of cigarette smoking on sperm parameters. J Int Med Res. 2010; 38(3): 923-928.
- Gaur DS, Talekar M, Pathak VP. Effect of cigarette smoking on semen quality of infertile men. Singapore Med J. 2007; 48(2): 119-123.
- Gholinezhad-Chari M, Hosseinzadeh-Colagar A. Seminal plasma lipid peroxidation, total antioxidant capacity, and cigarette smoking in asthenoteratospermic men. J Mens Health. 2011; 8(1): 43-49.
- 31. Pasqualotto FF, Sobreiro BP, Hallak J, Pasqualotto EB, Lucon AM. Cigarette smoking is related to a decrease in semen volume in a population of fertile men. BJU Int. 2006; 97(2): 324-326.
- Ramlau-Hansen CH, Thulstrup AM, Aggerholm AS, Jensen MS, Toft G, Bonde JP. Is smoking a risk factor for decreased semen quality?. A cross-sectional analysis. Hum Reprod. 2007; 22(1): 188-196.
- Mostafa T, Anis TH, El-Nashar A, Imam H, Othman IA. Varicocelectomy reduces reactive oxygen species levels and increases antioxidant activity of seminal plasma from infertile men with varicocele. Int J Androl. 2001; 24(5): 261-265.
- Pasupathi P, Saravanan G, Farook J. Oxidative stress bio markers and antioxidant status in cigarette smokers compared to nonsmokers. J Pharm Sci Res. 2009; 1(2): 55-62.
- Amiri I, Sheikh N, Najafi R. Nitric oxide level in seminal plasma and its relation with sperm DNA damages. Iran Biomed J. 2007; 11(4): 259-264.
- Huang I, Jones J, Khorram O. Human seminal plasma nitric oxide: correlation with sperm morphology and testosterone. Med Sci Monit. 2006; 12(3): CR103-106.
- Koppenol WH, Moreno JJ, Pryor WA, Ischiropoulos H, Beckman JS. Peroxynitrite, a cloaked oxidant formed by nitric oxide and superoxide. Chem Res Toxicol.

Seminal Oxidative Damage in Infertile Smoking Men

- 1992; 5(6): 834-842.
- 38. Hibbs JB Jr, Vavrin Z, Taintor RR. L-arginine is required for expression of the activated macrophage effector mechanism causing selective metabolic inhibition in target cells. J Immunol. 1987; 138(2): 550-565.
- Sheikh N, Amiri I, Najafi R, Goodarzi MT. The correlation between total antioxidant capacity and nitric oxide concentration in seminal plasma with sperm DNA damage. Afr J Biotechnol. 2010; 9(35): 5739-5745.