

Evaluation of The Number of CD4⁺ CD25⁺ FoxP3⁺ Treg Cells in Normal Mice Exposed to AFB₁ and Treated with Aged Garlic Extract

Mohaddeseh Larypoor, Ph.D.^{1*}, Mansour Bayat, Ph.D.¹, Zuhair Mohammad Hassan, Ph.D.²,
Abbas Akhavan Sepahy, Ph.D.³, Masoud Amanlou, Ph.D.⁴

1. Department of Mycology, Faculty of Veterinary, Science and Research Branch, Islamic Azad University, Tehran, Iran
2. Department of Immunology, School of Medical Sciences, Tarbiat Modares University, Tehran, Iran
3. Department of Microbiology, North Branch of Islamic Azad University, Tehran, Iran
4. Department of Medicinal Chemistry, Faculty of Pharmacy and Medicinal Plants Research Center, Tehran University of Medical Sciences, Tehran, Iran

* Corresponding Address: P.O.Box: 14515/775, Department of Mycology, Faculty of Veterinary, Science and Research Branch, Islamic Azad University, Tehran, Iran
Email: M_Larypoor@iauu-tmb.ac.ir

Received: 16/May/2012, Accepted: 11/Sep/2012

Abstract

Objective: Aflatoxin B₁ (AFB₁) suppresses the immune system. To decrease such suppressive effects on the immune system, a wide range of herbal medicines like garlic are utilized. Biological activities of garlic *in vitro* and *in vivo* have also been verified. Our previous studies demonstrated that aged garlic (dry garlic bulbs preserved in the freezer for six months at -20°C) have increased immunostimulator fractions and reduced immunosuppressor fractions. This study focuses on the immunosuppressor activity of AFB₁ and immunostimulator activity of aged garlic extract (AGE) through the evaluation of CD4⁺ CD25⁺ FoxP⁺ regulator cell (Treg) counts and the pattern of cytokine production in Balb/c normal mice.

Materials and Methods: In this experimental research, AFB₁ was separated from *Aspergillus flavus* (PTCC 5004) by HPLC and AGE prepared using the Mantis method. The Delayed-Type Hypersensitivity (DTH) test was carried out to determinate the effectiveness of different doses of AGE and AFB₁, which can both have an effect on the immune system. Subsequent experiments were carried out on 20 Balb/c mice to estimate the effects of AGE and AFB₁ on the number of Treg cell in 4 groups: 10 µl/kg/day of AFB₁ and AGE diluents were administered for 4 consecutive days to group 1. AFB₁, 2. control, 3. AGE + AFB₁ and 4. AGE via intraperitoneal (IP) route, respectively. Mice were sacrificed and splenocytes harvested and the percentage of splenic Treg cells was measured by flow cytometry analysis. The ELISA method was utilized to measure Cytokine production.

Results: The findings reveal that AGE increased the level of IFN-γ and IL-4 cytokines produced by splenocytes stimulated by specific tumor antigen and decreased the number of Treg cells in the spleen (p<0.05). AFB₁ increased the number Treg cells in the spleen and decreased cytokine production (p<0.05). In groups 2 (control) and 4 (AGE) the number of Treg cells decreased (p value<0.05) whereas in groups 1 and 3 the number of Treg cells increased (p<0.05).

Conclusion: This study indicated that AGE is able to alter the cytokine production in normal mice into a Th₁ protective pattern which is beneficial to the immune system in general and anti-tumor immunity in particular. AFB₁ is able to alter the cytokine production into a Th₂ protective pattern. Therefore, AGE might be used as herbal medicine with few side effects as compared to chemotherapy in treating cancers caused by substances like AFB₁.

Keywords: Aflatoxin-B₁, Garlic, Treg Cells, Immunotherapy

Cell Journal(Yakhteh), Vol 15, No 1, Spring 2013, Pages: 37-44

Citation: Larypoor M, Bayat M, Mohammad Hassan Z, Akhavan Sepahy A, Amanlou M. Evaluation of the number of CD4⁺ CD25⁺ foxP3⁺ Treg cells in normal mice exposed to AFB₁ and treated with aged garlic extract. cell J. 2013; 15(1): 37-44.

Introduction

AFB₁, a secondary metabolite of the fungus *Aspergillus flavus*, is a hepatocarcinogen in various animal species, including fish, birds, rodents, and nonhuman primates (1-4). It is also a suspected human carcinogen and has been shown to play a role in human hepatocarcinoma (5-7). When low dosages of AFB₁ are received on a daily basis over a long time, the number of Treg cells (CD4⁺ CD25⁺ FoxP3⁺) is changed in the human body (3,4). The authors present part of their immunotoxicity study, which aims to complement a larger, collaborative effort designed to assess potential biomarkers that may have a role in the initiation and promotional stages of carcinogenesis (9, 10), and which may be of relevance in the "risk assessment" process. In particular, the authors were interested in the effects of AFB₁ on cells and the mechanisms of cell-mediated immunity (CMI) as this has been implicated as the immune target (11, 12) with respect to carcinogenesis. AFB₁ has unique chemical structures, which cause harm by reacting with the chemicals in living organisms. The structure of AFB₁ is shown in figure 1.

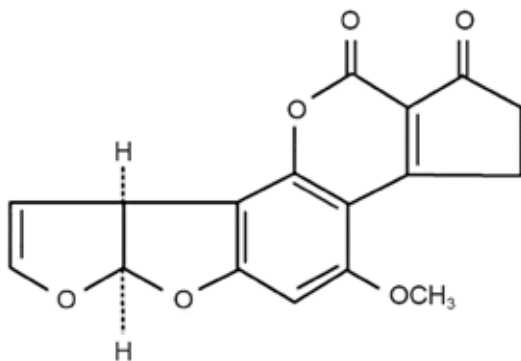


Fig 1: The structure of AFB₁.

As a digestive stimulant, diuretic, and antispasmodic, garlic (*Allium sativum*, Liliaceae) is used by many people all over the world. Garlic has recently been reported to have antibiotic

properties and benefits including antifungal (13) and antibacterial activities (14). It is also reported to have hypolipidemic, anti-atherosclerosis (15) and anti-carcinogenesis activities (16). Various research studies have indicated that garlic modulates immune responses (16, 17).

The authors' own previous studies have demonstrated that garlic enhances natural killer (NK) cell activity (17) and T-lymphocyte proliferation (18-20). Garlic extract and a garlic protein fraction were shown to augment the oxidative burst in peritoneal macrophages of Balb/c mice (21). Ghazanfari et al. showed that garlic extract induces a shift in cytokine pattern in Balb/c mice with a *Leishmania major* infection and an upshot in the immune response with regard to Th₁ (IFN- γ , IL-2) (22, 23). At the same time, a unique garlic preparation called AGE has been reported to have a series of pharmacologic effects including immunomodulation (20). In rodents, AGE and its constituents have been reported to inhibit the development of chemically-induced tumors in the bladder, mammary glands (24, 25), colon, esophagus, lung, skin and stomach (26). Recent studies have focused on the immunological behavior of AGE and AFB₁ (3). In the present study, the authors investigated the stimulation and suppression of the immune system of Balb/c mice *in vitro* by AFB₁ and AEG.

Materials and Methods

Materials

AFB₁ preparation

Toxicogenic *Aspergillus flavus* (PTCC 5004) was purchased from the Iranian Research Organization for Science and Technology (IRO-ST) and tested for the generation of AFB₁ by slide chromatography. The *Aspergillus flavus* was then cultured in Aflatoxin production medium to generate mycotoxin. AFB₁ separated of culture extract by HPLC method (1, 6).

Preparation of AGE

Fresh garlic bulbs were obtained from Hama-

dan, a city in western Iran and famous for its fresh garlic. Dry garlic bulbs were peeled and preserved in the freezer (-20°C) for six months. Aqueous aged garlic extract was prepared using the Mantis method (20). Garlic bulbs were homogenized with two parts of distilled water in a varying blender. The homogenized blend was filtered under vacuum through Whatman paper (No. 1) and the filtrate was centrifuged at 3400 g for 30 minutes. The clear supernatant was collected. Twenty-seven grams of NH_4SO_4 were added to one liter of the supernatant and centrifuged at 3400 g for 30 minutes. The residue was re-suspended in saline and dialyzed against buffer saline. AGE samples were then run on G 50 gel chromatography to measure protein using the Bradford assay and evaluated by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) (24).

SDS-PAGE electrophoresis

A 12% (weight/volume) polyacrylamide gel was utilized to judge the purity of molecules and to estimate the molecular mass compacted with proteins. After electrophoresis, the gel was fixated with methanol and acetic acid formaldehyde for 60 minutes and stained with coomassie blue.

The sample

The groups of inbred female Balb/c mice age 4-6 weeks were purchased from the Pasteur Institute of Iran. Four groups (five mice in each) were housed in a standard poly-propylene cage. The animals were kept under standard conditions (a cycle of 12/12 hour light/dark and a temperature of 20-22°C) with free access to water and autoclaved standard mouse chow. Animal care and treatment were conducted in conformity with the guidelines of Animal Care and Research Committee of Tarbiat Modares University and in compliance with the Guide for the Care and Use of Laboratory Animals [DHEW Publication No. (NIH) 85-23, Revised 1985, Office of Science and Health Reports, DRR/NIH, Bethesda, MD 20205].

Methods

Delayed-type hypersensitivity (DTH) test

To evaluate DTH response, 20 female mice

were randomly assigned into groups of five. On day 0, 0.1 ml of a solution containing 1×10^8 sheep red blood cells (sRBCs, Razi Institute, Tehran, Iran) suspended in PBS were subcutaneously injected in the back of all mice. Three groups received three doses (30, 15, 10 $\mu\text{l}/\text{kg}/\text{day}$) of AFB_1 (0.1 ml) via the intraperitoneal (IP) route over a 5-day period. The remaining group (the control group) received diluent [a solution of ethanol /PBS (40:60)] via the same route, dosage, and time interval. On the 5th day, the sensitized animals were subcutaneously challenged with 1×10^8 sRBCs in the left hind foot pads. The increase in the foot pad thickness was measured with a vernier calliper (Mitutoyo, Japan) one, two and three days after the booster injection of sRBCs. The results were calculated according to the following formula (18, 19):

$$\text{Increased DTH} = \frac{(\text{Left footpad challenged with sRBC} - \text{right footpad}) \times 100}{\text{Right footpad}}$$

Preparation of the animal model

The first group in this experimental study was treated with 0.1 ml of AFB_1 at a daily dose of 10 $\mu\text{l}/\text{kg}$ via the IP route. This dose had already been defined by the DTH test (above) as the optimal immunostimulatory dose. The second group (negative control) also received the same volume of diluents PBS via the IP route. The third group was treated with 0.1 ml of AFB_1 (10 $\mu\text{l}/\text{kg}/\text{day}$) and 0.1 ml of AGE (20 mg/kg/day) via the IP route (25) and the fourth group with 0.1 ml of AGE (20 mg/kg/day) via the same route. The treatments were applied once in every day in 7 day period (26).

Splenocyte cytokine production measurement through ELISA method

The isolated spleen mononuclear cells were cultured in 24-well plates (Nunc, Denmark) in a final concentration of 2×10^6 cells/ml. Three samples were taken from each mouse in the group and each sample was analyzed in triplicate. Twenty microliters of purified tumor antigen and phytohemagglutinin (PHA) were added to each separately to stimulate the cells. After

three day incubation at 37°C and 5% CO₂, the supernatants were collected and frozen at -70°C until analyzed by enzyme-linked immunosorbent measurement (ELISA). IFN- γ and IL-4 concentrations were measured using the R&D American DuoSet ELISA Development kit. The treatment mice were unconscious and medullary and separated spleen.

Separation of splenic mononuclear cells (MNC)

The control and treated animals were sacrificed by cervical dislocation on the 13th day; spleens were resected under sterile conditions and were suspended in PBS. The splenic cell suspension was RBC- lysed with 0.75% NH₄Cl and Tris buffer (0.02%) (pH=7.4). The cells were washed and the single-cell suspension was prepared in RPMI-1640 (Gibco, 51800-035, Stey cell Technology Company) containing stable glutamine (Cytogen, USA) and 10% heat inactivated fetal calf serum (Gibco, England). To define the viability and density of cells in the suspension the Trypan blue dye exclusion method was used. The cells were counted using homocytometer light microscopy. The viability of the splenocytes was generally above 95%. After an additional washing, the suspension was adjusted to 4 \times 10⁶ cells per milliliters in RPMI-1640 supplemented with 10% FCS, 100 μ g/mL streptomycin, and 100 IU/mL penicillin (complete RPMI), and kept at 4°C.

Three-color immunostaining and flow cytometry analysis

After treating the mice during the 7-day period as mentioned in 2.2, the MNCs purified from the mice spleens were immunostained with the FITC anti-mouse CD4, PE-Cy5 anti-mouse CD25 (BD, eBioscience, USA), and subsequently with PE-Cy5 anti-mouse FoxP3⁺, according to the eBioscience mouse regulatory Tcell staining kit's instruction. Three samples were taken from each mouse and each was analyzed in triplicate. The samples were analyzed using a FACSCalibur flow cytometer at Tehran University and the results were analyzed with WinMDI/25 software.

Statistical analyses

In this study each experiment was performed in duplicate or triplicate and one-way analysis of var-

iance (ANOVA) or Mann-Whitney non-parametric tests were used to determine the statistical significance ($p < 0.05$) between values in the experimental and control groups. The data were analyzed using SPSS software version 16 and the results are expressed as measures of central tendency and dispersion (mean, SE, etc.).

Results

Bradford assay and SDS-PAGE electrophoresis

The results of the Bradford evaluation showed that the quantity of the effective protein in AGE is 0.27 μ g/ml. Gel electrophoresis was performed. The results are shown in figure 2.

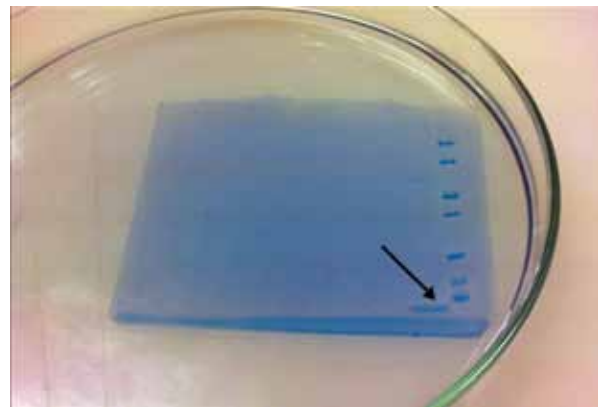


Fig 2: Gel electrophoresis: protein bound in AGE is shown by arrow-head.

Effect of AFB₁ on DTH test

In order to estimate the effect of AFB₁ on cell mediated immunity (CMI), twenty mice (four groups of five mice) were treated with AFB₁ and PBS as shown in figure 3. For five consecutive days, the mice were sensitized using sRBCs, treated with three doses of AFB₁ (30, 10, 5 μ l/Kg/Day in three groups, and PBS: ethanol 60:40 in the control group). A challenge using sRBCs was then performed in the left foot pad.

The percentage of foot pad swelling was measured using a digital vernier capillier at intervals of one, two and three days. There was a significant difference in mice treated with a dose

of 10 $\mu\text{g}/\text{kg}/\text{day}$ compared to control mice two and three days after the foot pad challenge. The steady increase in the pad swelling two and three days after injection (p value=0.01) showed that a dose of 10 $\mu\text{l}/\text{kg}/\text{day}$ of AFB_1 significantly contributed to a greater DTH response every one day after the foot pad challenge compared to controls ($p=0.01$, 0.046 and 0.021 respectively). This increase was not seen in other groups. As a consequence, the optimum dose of 10 $\mu\text{l}/\text{kg}/\text{day}$ of AFB_1 was used for the rest of the investigations.

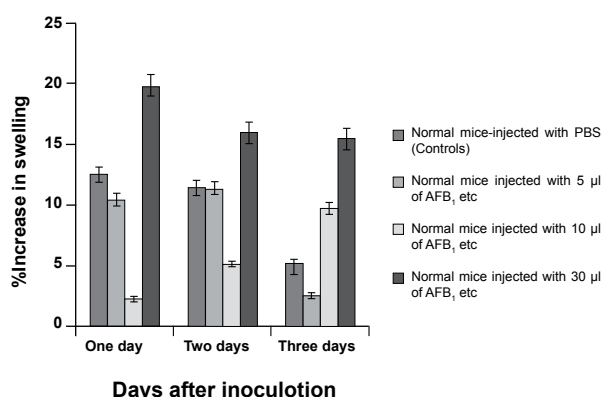


Fig 3: The results of delayed type hypersensitivity (DTH) assay. As shown in the graph a significant difference in the degree of swelling was detected in the group treated with 10 μl of AFB_1 ($p=0.015$ and 0.021 at two and three respectively).

Effect of AFB_1 and AGE on splenic $\text{CD4}^+ \text{CD25}^+ \text{FoxP3}^+$ T cells

Concentration of $\text{IFN-}\gamma$ and IL-4 , typical cytokines for Th_1 and Th_2 (Previously you described Th_1 as follows: Th_1 ($\text{IFN-}\gamma$, IL-2) and Th_2 pattern has not been explained at all.) pattern, in treated and untreated mice was evaluated by ELISA technique. The results demonstrated that mice treated with AFB_1 showed a decreased level of $\text{IFN-}\gamma$ and an increased level of IL-4 , but in the case of mice treated with AGE, the results showed an increased level of $\text{IFN-}\gamma$ and a decreased level of IL-4 . These differences were statistically significant ($p<0.05$). The results for the $\text{IFN-}\gamma$ and IL-4 concentra-

tions are shown in figure 4 and figure 5.

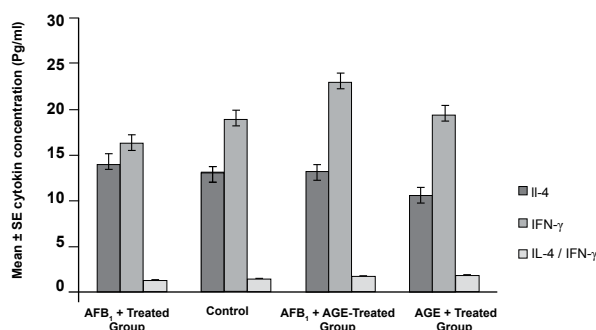


Fig 4: Results of ELISA assessment showing the level of $\text{IFN-}\gamma$ and IL-4 cytokines produced from splenocytes stimulated by specific tumor antigen. Results show a statistically significant difference between the AGE-treated group and AFB_1 -treated group ($p<0.05$).

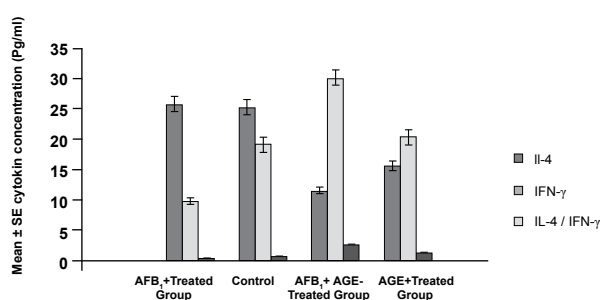


Fig 5: Results of ELISA assessment showing the level of $\text{IFN-}\gamma$ and IL-4 cytokines produced from splenocytes stimulated by specific PHA. Results show a statistically significant difference between the AGE-treated group and AFB_1 -treated group ($p<0.05$).

Effect of AFB_1 and AGE on splenic $\text{CD4}^+ \text{CD25}^+ \text{FoxP3}^+$ T cells

The flow cytometry technique was used to define the percentage of splenic Treg in Balb/c mice. As shown in figure 6 (A and B) the results indicate a statistically significant difference between the percentage of splenic Treg cells in the AGE-treated group and the AFB_1 -treated group and the control group and the AFB_1 + AGE-treated group. The percentage of splenic Treg cells in the AGE-treated group was lower than the AFB_1 -treated group ($p=0.049$).

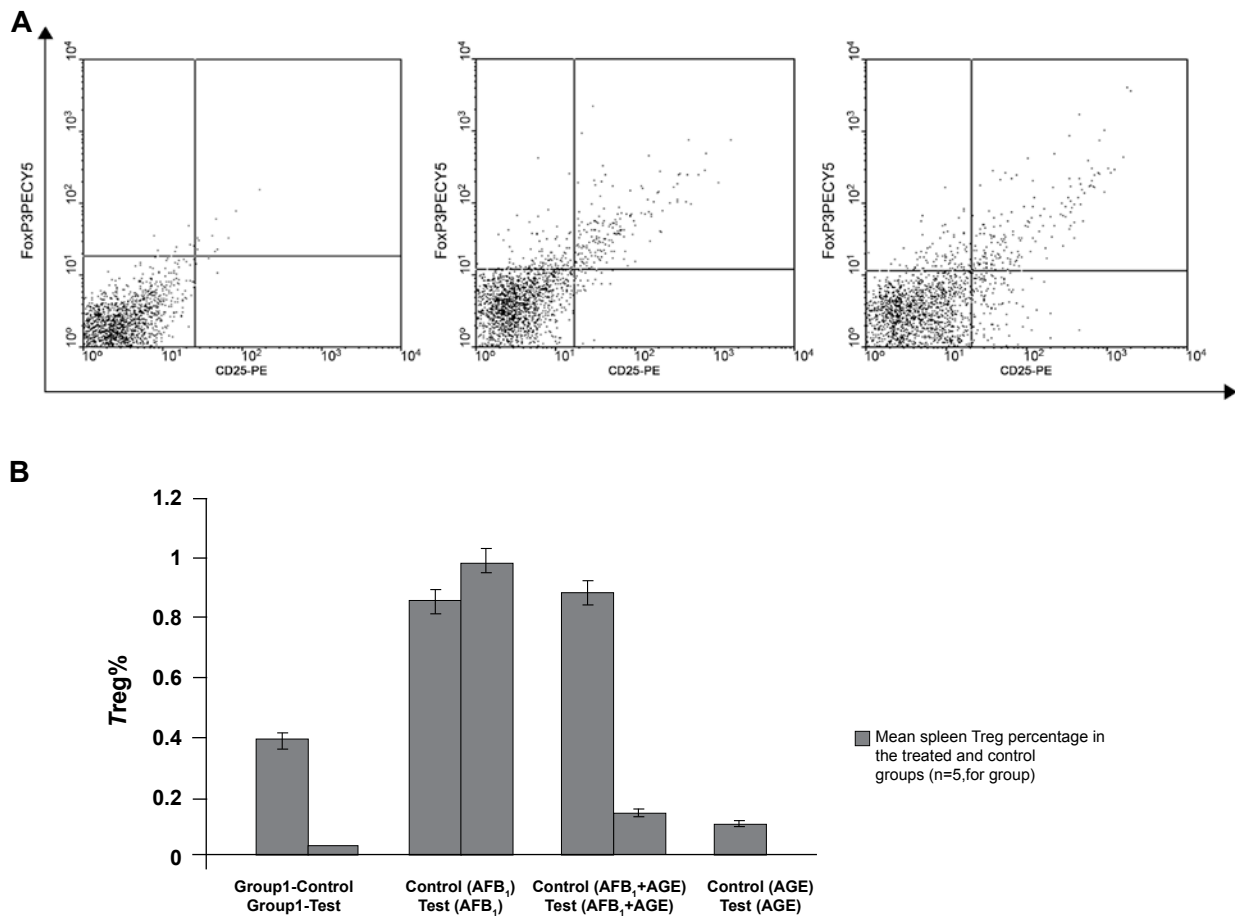


Fig 6: Results of flow cytometry measurement: A. Dot plots representing mean percentage of CD25⁺ and Foxp3⁺ expressing cells on the CD₄⁺ lymphocyte gate of splenocytes. B. Mean \pm SE spleen %Treg cells in treated and control groups. (p value=0.049).

Discussion

AFB₁, a secondary metabolite of *Aspergillus flavus*, has unique chemical structures. These structures react and interact with the chemicals in living organisms causing harm. *Aspergillus flavus* is a hepatocarcinogen in humans and can invade tumor cells. If a human receives a small dose of AFB₁ on daily basis over a long period of time, it will result in carcinoma (1, 5-7). Unfortunately, it is possible for humans to receive small doses of AFB₁ while consuming contaminated food, specifically milk. AFB₁ effects on the immune system can be either stimulatory or suppressive depending on the critical exposure windows of dose and time (9, 10). The immune cells in the spleen, such as T-lymphocytes and macrophages, both significant mediators of inflammatory responses to tissue damage, have

been shown to be differently affected by continuous and intermittent exposures to AFB₁ (11, 12).

During the past decade, medical researchers have increasingly focused on herbal medicine, especially Garlic. Garlic has been consumed for food and medicinal purposes worldwide for thousands of years. Garlic's beneficial effects on human health are known to everyone (22, 24). Currently, the garlic plant itself, as well as its numerous extracts, are commercially available as dietary supplements (20, 22, 24). Epidemiological studies suggest garlic consumption has preventive effects in some types of cancer (18). Various researchers have indicated that garlic modulates immune responses (17, 18). Previous studies showed that garlic enhances natural killer cell (NK) cell activity and T-lymphocyte proliferation (19). Also garlic extract and a gar-

lic protein fraction have been shown to augment the oxidative burst in peritoneal macrophages of Balb/c mice (26). Lau et al. showed that AGE is an efficient candidate as an immune modifier compared to fresh garlic extract, which maintains the homeostasis of immune functions (22). In the present investigation, the authors explored the cytotoxicity and immunomodulatory activities of AFB₁ and AGE *in vivo* in a mouse model. First, in order to select the optimal immunostimulatory dose of AFB₁, the researchers performed DTH measurements. The results demonstrated that a dose of 10 µl/Kg/Day had a significant effect on the DTH test two and three after a foot pad challenge. Based on previous research a dose of 20 mg/kg/Day of AGE were used to perform these tests (17, 18, 20). Our own findings and those of other studies (11, 24-25) found a dose of 10 µl/Kg/Day to be the optimal immunostimulatory dose and this dose was used in all further experimentation. Nevertheless, our results showed that AGE have effects of supporting of immune system. Figure 5 and figure 6 indicate that in AFB₁-treated groups the level of IFN-γ cytokines decreased in the control group whereas in the AGE-treated groups the level of IFN-γ cytokines increased. Increased production of IFN-γ and decreased IL-4 in turn have other anti-cancer effects including anti-angiogenesis, increased NK activity and increased immune activity. Therefore it seems that a continuous cascade of cytokine production and cellular activation, by definition, explain the AGE anti-cancer mechanisms and increased activation of the immune system. This finding was confirmed in a previous study by Hassan et al. (22) and Noori et al. (23) which indicated that AGE is able to enhance the capacity of splenic leukocytes to produce IFN-γ and IL-4 following mitogenic stimulation. The authors also showed that AGE could decrease and that AFB₁ could increase the total number of splenic Treg cells in the experimental group. Although the mechanism for Treg reduction by AGE is not yet fully understood, given that AGE has been shown to decrease tumor growth (24), decrease in the number of Treg cells may result in tumor size reduction. Moreover, some studies have reported that AGE can reduce the number of Treg cells via the inhibition of NO production "an inducer metabolite for Treg expansion" from macrophage cells (21). This finding is confirmed in a previous study by Noori et al. (23) which showed that AGE is able to reduce Treg and

inhibit tumor growth *in vivo* but AFB₁ is able to increase Treg and stimulation tumor growth *in vivo*.

Conclusion

Overall, based on the findings of this research and other studies, the authors measured and showed the specific immunomodulatory properties of AFB₁ that are needed to suppress the immune system and the specific immunomodulatory properties of AGE that are needed to support the immune system. Immune cells in spleen such as T-lymphocytes and macrophages, important mediators of inflammatory responses to tissue damage and cancer, were affected differently by continuous and intermittent exposure to AFB₁. This study showed that AGE decreases the production of Treg cells from splenocytes. Taken together, the findings suggest that AGE may play a role in attenuating tumor growth by increasing cytokine production and decreasing Treg cells and anti-tumoral cell activation during cell carcinogenesis. It is possible to speculate that AGE could be used as a plant drug with few side effects during anticancer chemotherapy.

Acknowledgments

The present study received a grant from the Parsroos Company. Immunological tests for this research were performed at the Department of Immunology at Tarbiat Modarres University of Medical Sciences. The authors wish to express their sincere appreciation for excellent technical assistance from Mr. Mahdavi and Miss Langroodi in the immunological analysis, Mr. Tebyanian in flow cytometry and Dr. Mostaffai in helping the authors with statistical analyses of the obtained data. There is no conflict of interest in this article.

References

1. Hinton DM, Myers MJ, Raybourne RA, Francke-Carroll S, Sotomayor RE, Shaddock J, et al. Immunotoxicity of aflatoxin B1 in rats: effects on lymphocytes and the inflammatory response in a chronic intermittent dosing study. *Toxicol Sci.* 2003; 73(2): 362-377.
2. Wogan GN. Impacts of chemicals on liver cancer risk. *Semin Cancer Biol.* 2000; 10(3): 201-210.
3. Wogan GN. Aflatoxins as risk factors for hepatocellular carcinoma in humans. *Cancer Res.* 1992; 52(7 Suppl): 2114s-2118s.
4. Jiang Yi, Pauline EJ, William OE, Wang Js, Timothy DP. Aflatoxin B1 albumin adduct levels and cellular immune status in Ghanaians. *Int Immunol.* 2005; 17(6): 807-814.
5. Hochstenbach K, van Leeuwen DM, Gmuender H, Stølevik SB, Nygaard UC, Løvik M, et al. Transcriptomic profile indicative of immunotoxic exposure: *in vitro* studies

- in peripheral blood mononuclear cells. *Toxicol Sci.* 2010; 118(1): 19-30.
6. Dimitri RA, Gabal MA. Immunosuppressant activity of aflatoxin ingestion in rabbits measured by response to *Mycobacterium bovis* antigen I. Cell mediated immune response measured by skin test reaction. *Vet Hum Toxicol.* 1996; 38(5): 333-336.
 7. Moon Ey, Rhee DK, Pyo S. Inhibition of various functions in murine peritoneal macrophages by aflatoxin B1 exposure in vivo. *Int J Immunopharmacol.* 1999; 21(1): 47-58.
 8. Jiang Y, Jolly PE, Ellis WO, Wang JS, Phillips TD, Williams JH. Aflatoxin B1 albumin adduct levels and cellular immune status in Ghanaians. *Int Immunol.* 2005; 17(6): 807-814.
 9. Moore GS, Atkins RD. The fungicidal and fungistatic effects of an aqueous garlic extract on medically important yeast-like fungi. *Mycologia.* 1977; 69(2): 341-348.
 10. Chung JG, Chen GW, Wu LT, Chang HL, Lin JG, Yeh CC, et al. Effects of garlic compounds diallyl sulfide and diallyl disulfide on arylamine N-acetyltransferase activity in strains of *Helicobacter pylori* from peptic ulcer patients. *Am J Clin Nutr Med.* 1998; 26(3-4): 353-364.
 11. Kritchevsky D. The effect of dietary garlic on the development of cardiovascular disease. *Trends Food Sci Technol.* 1991; 2(6): 141-144.
 12. Yaraei R, Saraf-Nejad A, Nozari B, Ghazanfari T, Hassan ZM. The effect of garlic extract and its fractions on natural killer activity. Abstract Book of 10th International Congress of Immunology, India: Bruce Robinson Publications; 1998: 493.
 13. Ghazanfari T, Hassan ZM. A lectine with mitogenic activity in garlic. Abstract Book of 10th International Congress of Immunology, India: Bruce Robinson Publications; 1998: 493.
 14. Lau BH, Yamasaki T, Gridley DS. Garlic compounds modulate macrophage and T-lymphocyte functions. *Mol Biother.* 1991; 3(2): 103-107.
 15. Ghazanfari T, Hassan ZM, Ebtekar M, Ahmadiani A, Naderi G, Azar A. Garlic induces a shift in cytokine pattern in *Leishmania major*-infected BALB/c mice. *Scand J Immunol.* 2000; 52(5): 491-496.
 16. Abdullah TH, Kirkpatrick DV, Carter J. Enhancement of natural killer cell activity in AIDS with garlic. *Dtsch Z Onkol.* 1989; 21: 52-53.
 17. Amagase H, Milner JA. Impact of various sources of garlic and their constituents on 7, 12-dimethylbenz[a]anthracene binding to mammary cell DNA. *Carcinogenesis.* 1993; 14(8): 1627-1631.
 18. Wattenberg LW, Sporn VL, Barany G. Inhibition of N-nitrosodiethylamine carcinogenesis in mice by naturally occurring organosulfur compounds and monoterpenes. *Cancer Res.* 1989; 49(10): 2689-2692.
 19. Ghazanfari T, Hassan ZM, Ebrahimi M. Immunomodulatory activity of a protein isolated from garlic extract on delayed type hypersensitivity. *Int J Immunopharmacol.* 2002; 2(11): 1541-1549.
 20. Ahmadabad HN, Hassan ZM, Safari E, Bozorgmehr M, Ghazanfari T, Moazzeni SM. Evaluation of the immunomodulatory effect of the 14 kDa protein isolated from aged garlic extract on dendritic cells. *Cell Immunol.* 2011; 269(2): 90-95.
 21. Nikoo S, Bozorgmehr M, Namdar Ahmadabad H, Hassan ZM, Moazzeni SM, Pourpak Z, et al. The 14kDa protein molecule isolated from garlic suppresses indoleamine 2, 3-dioxygenase metabolites in mononuclear cells in vitro. *Iran J Allergy Asthma Immunol.* 2008; 7(4): 203-208.
 22. Hassan ZM, Yaraee R, Zare N, Ghazanfari T, Sarraf Nejad AH, Nazori B. Immunomodulatory affect of R10 fraction of garlic extract on natural killer activity. *Int Immunopharmacol.* 2003; 3(10-11): 1483-1439.
 23. Noori S, Taghikhani M, Hassan ZM, Allameha A, Mostafaei A. Tehranolide molecule modulates the immune response, reduce regulatory T cell and inhibits tumor growth in vivo. *Mol Immunol.* 2010; 47(7-8): 1579-1584.
 24. Farsam V, Hassan ZM, Zavarani-Hosseini A, Noori S, Mahdavi M, Ranjbar M. Antitumor and immunomodulatory properties of artemether and its ability to reduce CD4⁺ CD25⁺ FoxP3⁺ T reg cells in vivo. *Int Immunopharmacol.* 2011; 11(11): 1802-1808.
 25. Theumer MG, López AG, Masih DT, Chulze SN, Rubinstein HR. Immunobiological effects of AFB₁ and AFB₁-FB1 mixture in experimental subchronic mycotoxicoses in rats. *Toxicology.* 2003; 186(1-2): 159-170.
 26. Dugyala RR, Sharma RP. The effect of aflatoxin B1 on cytokine mRNA and corresponding protein levels in peritoneal macrophages and splenic lymphocytes. *Int J Immunopharmacol.* 1996; 18(10): 599-608.